Lalević, B., Gavrić, T., Stambolić, A., Sunulahpašić, A., Komlen, V., Čengić, L., Hamidović, S. (2024). Antifungal potential of peppermint, basil and sage essential oils. Agriculture and Forestry, 70 (3): 115-121. https://doi.org/10.17707/AgricultForest.70.3.08

DOI: 10.17707/AgricultForest.70.3.08

Blažo LALEVIĆ^{1*}, Teofil GAVRIĆ², Amina STAMBOLIĆ², Amer SUNULAHPAŠIĆ³, Vedrana KOMLEN⁴, Lejla ČENGIĆ², Saud HAMIDOVIĆ²

ANTIFUNGAL POTENTIAL OF PEPPERMINT, BASIL AND SAGE ESSENTIAL OILS

SUMMARY

Synthetic fungicides are the most effective protection against plant pathogens, but, their uncontrolled and long-term use can lead to many harmful effects: environmental degradation, human health problems and pathogen resistance. The biological compounds contained in essential oils have no harmful effects on humans or the environment and can therefore be an alternative to synthetic fungicides. Essential oils are products of plant metabolism and often show antifungal, antiviral, antibacterial and insecticidal effects. The aim of this study was to investigate the antimicrobial potential of essential oils from peppermint, basil and sage essential oils obtained by hydrodistillation, on Fusarium sp. and Aspergillus sp. The experiment was carried out on potato dextrose agar. After inoculation of the agar with fungal mycelia, paper discs impregnated with 10 μ l of oils were placed on the agar surface. In the control, the impregnation was carried out with distilled water. The inhibition zones were measured after 3, 6 and 9 days. The results showed that peppermint oil had the highest antimicrobial potential compared with other oils. Sage essential oil showed the lowest antifungal suppression. A negligible zone of inhibition was observed in the control. A statistically significant influence between the oils and the incubation period was found in this study. Our results confirm the potential use of peppermint essential oil for the suppression of *Fusarium* sp. and *Aspergillus* sp. growth.

Keywords: antifungal potential, *Aspergillus*, *Fusarium*, inhibition zone, plant essential oils.

¹ Blažo Lalević^{*} (corresponding author email: blazol@agrif.bg.ac.rs), University of Belgrade, Faculty of Agriculture, Nemanjina 6, Belgrade-Zemun, SERBIA

² Teofil Gavrić, Amina Stambolić, Lejla Čengić, Saud Hamidović, University of Sarajevo, Faculty of agriculture and food science, Zmaja od Bosne 8, Sarajevo, BOSNIA AND HERZEGOVINA;

³ Amer Sunulahpašić, Ministry of agriculture, water management and forestry, Travnik, BOSNIA AND HERZEGOVINA

⁴ Vedrana Komlen, The University "Džemal Bijedić", Agromediterranean Faculty, University Campus, Mostar, BOSNIA AND HERZEGOVINA

Notes: The authors declare that they have no conflicts of interest. Authorship Form signed online. Received:07/03/2024 Accepted:18/07/2024

INTRODUCTION

Agricultural plant production is often associated with phytopathogen infestation (Köhl et al., 2019) and yield losses (Ampt et al., 2019). Chatterjee et al. (2016) found that the occurrence of plant disease leads to an annual loss of 10-15 percent of major crops worldwide. Peng et al. (2021) point out that more than twothirds of plant diseases are associated with the occurrence of pathogenic fungi. Therefore, plant diseases need to be controlled using different techniques (Tsegaye et al., 2018), which requires redesigning agroecosystems to improve plant resistance to pathogens (McDonald and Stukenbrock, 2016). Pesticides show high efficiency in suppressing the occurence of Fusarium (Arie, 2019) and Aspergillus (Geetha et al., 2016); species of these fungal genera are frequently detected on various plant materials (Kalman et al., 2020; Perrone et al., 2007). However, Aktar et al. (2009) demonstrated that the use of pesticides is associated with higher plant productivity, avoidance of yield losses and control of disease vectors. On the other hand, the extensive use of pesticides led to significant degradation of the agroecosystem and human health (Salameh et al., 2006; Zheng et al., 2016). Therefore, several alternative methods to replace pesticides have been proposed (Nile et al., 2019).

The pesticidal properties of many plant species and their products are wellknown and may represent formulations with potential applications against pathogens (Stevenson *et al.*, 2017). Nxumalo *et al.* (2021) described the possibility of medicinal plant extracts as an alternative technology for synthetic chemicals. Several *in vitro* studies have been conducted to determine the potential of various plant extracts in suppressing growth of fungal genera such as *Candida*, *Aspergillus*, *Penicillium*, etc. (Azaz *et al.*, 2004; Chuang *et al.*, 2007; Hammer *et al.*, 1999; Omran and Esmailzadeh, 2009).

The objective of this paper is to evaluate the antifungal potential of the essential oils of peppermint, basil and sage against *Fusarium* sp. and *Aspergillus* sp. in *in vitro* experiment.

MATERIAL AND METHODS

Fusarium sp. and *Aspergillus* sp. used in this research belong to the collection of microorganisms Department of Microbiology at the Faculty of agriculture and food science in Sarajevo (Bosnia and Herzegovina). These fungal isolates were previously cultivated and stored at 4 °C on the nutrient medium potato dextrose agar (PDA).

The shoots of peppermint, basil and sage were used for the extraction of plant material. Extracts were obtained through hydrodistillation (30 g of plant material and 400 ml of distilled water) using Neo Clevenger-type apparatus. The antifungal properties of plant extracts were determined using a disc diffusion assay. Potato dextrose agar (Himedia, India), previously sterilized at 120 °C for 20 min, was inoculated with pure cultures of *Fusarium* sp. and *Aspergillus* sp. Fungal mycelia were placed at four places on the agar surface, about 5 mm from the edge of the Petri dishes. In the center of the agar plate, one sterile filter paper disc (about

5 mm in diameter) containing 10 μ l of the plant extract was placed. In control, distilled water was used for the impregnation of the disc. All experiments were performed in triplicate. Incubation of Petri dishes was performed at 22 °C for 9 days. All experiments were performed in 4 replications. The zone of inhibition (expressed in centimeters) was measured on the 3, 6 and 9 day of incubation.

For the determination of the statistical significance of obtained results, SPSS software (version 22, SPSS Inc., Chicago, IL) was used. P<0.05 was chosen as a parameter of statistical significance between the treatments.

RESULTS AND DISCUSSION

Our results showed that all tested essential oils had an inhibitory effect on the growth of *Fusarium* sp. (Table 1). The degree of inhibition depended on the essential oil and the incubation period.

All tested oils showed significantly higher values of the inhibition zone compared to the control (Table 1). After three, six and nine days of incubation, peppermint essential oil showed the statistically highest effect. The diameter of the inhibition zone with peppermint essential oil was 1.58, 0.68 and 0.23 cm, respectively. Sage essential oil showed a larger diameter of the inhibition zone after three days, while after six and nine days, the effect of basil essential oil was more pronounced compared with sage. In all treatments, the diameter of the inhibition zone statistic decreased significantly with increasing of incubation time for peppermint and sage essential oils. In the control, no zone of inhibition was observed after 6 and 9 days.

D	Essenti	ial oils										
	peppermint			basil		sage			control			
	Time of incubation (days)											
	3	6	9	3	6	9	3	6	9	3	6	9
1	1.6	0.7	0.2	0.8	0.4	0.2	0.8	0.3	0.0	0.3	0.0	0.0
2	1.5	0.6	0.2	0.6	0.2	0.0	0.8	0.3	0.0	0.3	0.0	0.0
3	1.3	0.5	0.2	0.5	0.2	0.0	0.8	0.3	0.1	0.2	0.0	0.0
4	1.9	0.9	0.3	0.8	0.3	0.1	0.7	0.2	0.0	0.2	0.0	0.0
Α	1.58ª	0.68ª	0.23°	0.68ª	0.28 ^c	0.75°	0.78^{a}	0.28 ^b	0.03°	0.25ª	0.0^{b}	0.0 ^b
Т	0.83ª			0.41 ^b			0.36 ^c			0.08 ^d		
	Legend: $D - discs; A - average; T - total average$											

Table 1. Diameter of inhibition zone (cm) of *Fusarium* sp.

Legend: D – discs; A – average; T – total average

The essential oils of peppermint, basil and sage had an inhibitory effect on the growth of *Aspergillus* sp. (Table 2). Significantly lower values for the diameter of the inhibition zone diameter were observed for all essential oils during incubation. As in the experiment with *Fusarium* mycelia, peppermint essential oil showed the highest zone of inhibition compared with other treatments (0.63, 0.35 and 0.15 cm after 3, 6, and 9 days of incubation, respectively). Basil essential oil showed a stronger effect than sage essential oil, especially after three and six days. At the end of the incubation period, the same values were found for the diameter of inhibition zone. In the control, a negligible zone of inhibition was observed after three days; further incubation revealed no zone of inhibition.

D	Essential oils								Control			
	peppermint			basil			sage)				
		Time of incubation (days)										
	3	6	9	3	6	9	3	6	9	3	6	9
1	0.7	0.4	0.2	0.6	0.3	0.1	0.4	0.2	0.1	0.1	0.0	0.0
2	0.7	0.4	0.2	0.5	0.2	0.0	0.3	0.1	0.0	0.0	0.0	0.0
3	0.5	0.3	0.1	0.4	0.2	0.0	0.4	0.2	0.0	0.0	0.0	0.0
4	0.6	0.3	0.1	0.5	0.2	0.0	0.2	0.1	0.0	0.0	0.0	0.0
Α	0.63ª	0.35 ^b	0.15°	0.5ª	0.23 ^b	0.03 ^c	0.33ª	0.15 ^b	0.03°	0.03ª	0.0^{b}	0.0^{b}
Т	0.38ª			0.25 ^b			0.17°			0.01 ^d		

Table 2. Diameter of inhibition zone (cm) of Aspergillus sp.

Legend: D – discs; A – average; T – total average

Essential oils from medicinal and aromatic plants have antimicrobial properties and can be used to combat plant pathogens. Essential oils from the *Lamiaceae* family are known as antifungal agents (Santra and Banarjee, 2020). Plants from the *Lamiaceae* family are distributed worldwide and represent a cost-effective source for the extraction of essential oils that can be used in agriculture (Feng *et al.*, 2011; Mamgain *et al.*, 2013). Essential oils from *Lamiaceae* are often used to control fungal pathogens in crop production, such as *Fusarium*, or fungi responsible for food spoilage, such as *Aspergillus* (Couladis *et al.*, 2004; Soliman and Badeaa, 2002).

Peppermint essential oil was recommended as the best inhibitor of *Fusarium* growth compared with other natural products (Kumar *et al.*, 2016). Helal *et al.* (2006) found that the application of peppermint essential oil was effective against 11 fungi, including *Aspergillus* species. Our results are consistent with the report of Guynot *et al.* (2003) who found that peppermint essential oil inhibited the growth of *Aspergillus* and *A. niger*.

Kocić-Tanaskov *et al.* (2011) reported the inhibition of *Fusarium* species growth using different concentrations of basil essential oil. The highest inhibitory effect of basil oil against *Fusarium* growth was described by Hashem *et al.* (2010). Our results are in agreement with those of Pandey and Dubey (1994), who described moderate to poor antifungal activity of basil essential oil.

Our results showed that the lowest inhibition zone of fungal growth was achieved with sage essential oil. Ferdes *et al.* (2017) described the low effect of sage essential oil on the growth of *A. niger* and *Fusarium oxysporum*. Sage essential oil showed no significant suppression of the growth of *Aspergillus flavus* (Foltinova *et al.*, 2017). However, the fungotoxic effect of sage essential oil was observed at higher concentrations of this product (Daferera *et al.*, 2003).

CONCLUSION

This study confirms that the presented essential oils can be used to control the growth of *Fusarium* and *Aspergillus* mycelia. The highest inhibition zone was observed with the use of peppermint and the lowest with the use of sage essential oil. Further studies will focus on the inhibition of other fungal species and a detailed content analysis of these natural plant products.

REFERENCES

- Aktar, Md.W., Sengupta, D., & Chowdhury, A. (2009): Impact of pesticides use in agriculture: their benefits and hazards. *Interdiscip. Toxicol.*, 2: 1–12.
- Ampt, E.A., van Ruijven, J., Raaijmakers, J.M., Termorshuizen, A.J., & Mommer, L. (2019): Linking ecology and plant pathology to unravel the importance of soil-borne fungal pathogens in species-rich grasslands. *Eur. J. Plant Pathol.*, 154: 141–156.
- Arie T. (2019): *Fusarium* diseases of cultivated plants, control, diagnosis, and molecular and genetic studies. *J. Pestic. Sci.*, 44: 275–281.
- Azaz, A.D., Irtem, H.A., Kurkcuoğlu, M., & Baser, K.H.C. (2004): Composition and the in vitro antimicrobial activities of the essential oils of some Thymus species. Z. *Naturforsch. C*, 59: 75–80.
- Chatterjee, S., Kuang, Y., Splivallo, R., Chatterjee, P, & Karlovsky, P. (2016): Interactions among filamentous fungi Aspergillus niger, Fusarium verticillioides and Clonostachys rosea: fungal biomass, diversity of secreted metabolites and fumonisin production. BMC Microbiol., 16: 83.
- Chuang, P.H., Lee, C.W., Chou, J.Y., Murugan, M., Shieh, B.J., & Chen, H.M. (2007): Anti-fungal activity of crude extracts and essential oil of Moringa oleifera Lam. *Bioresour. Technol.*, 98: 232–236.
- Couladis, M., Tzakou, O., Kujundzic, S., Sokovic, M., & Mimica-Dukic, N. (2004): Chemical analysis and antifungal activity of *Thymus striatus*. *Phytother. Res.*, 18: 40–42.
- Daferera, D.J., Ziogas, B.N., & Polissiou, M.G. (2003): The effectiveness of plant essential oils on the growth of *Botrytis cinerea*, *Fusarium* sp. and *Clavibacter michiganensis* subsp. *Michiganensis*. Crop Prot., 22: 39–44.
- Deans, S.G., Svoboda, K.P., Gundidza, M., & Brechany, E.Y. (1992): Essential oil profiles of several temprate and tropical aromatic plants: their antimicrobial and antioxidant activities. *Acta Hortic.*, 306: 229–233.
- Dwivedi, S.L., Lammerts van Bueren, E.T., Ceccarelli, S., Grando, S., Upadhyaya, H.D., & Ortiz, R. (2017): Diversifying food systems in the pursuit of sustainable food production and healthy diets. *Trends Plant Sci.*, 22: 842–856.
- Feng, W., Chen, J., Zheng, X., & Liu, Q. (2011): Thyme oil to control Alternaria alternata in vitro and in vivo as fumigant and contact treatments. *Food Control*, 22: 78–81.
- Ferdes, M., Juhaimi, F.A., Özcan, M.M., & Ghafoo, K. (2017): Inhibitory effect of some plant essential oils on growth of Aspergillus niger, Aspergillus oryzae, Mucor pusillus and Fusarium oxysporum. S. Afr. J. Bot., 113: 457–460.
- Foltinova, D., Tančinová, D., & Císarová, M. (2017): Influence of essential oils on the growth of *Aspergillus flavus*. *Slovak J. Food Sci.*, 11: 322–331.
- Godfray, H.C.J. (2014): The challenge of feeding 9–10 billion people equitably and sustainably. J. Agric. Sci., 152: 2–8.

- Guynot, M.E., Ramos, A.J., Seto, L., Purroy, P., Sanchis, V., & Marin, S. (2003): Antifungal activity of volatile compounds generated by essential oils against fungi commonly causing deterioration of bakery products. J. Appl. Microbiol., 94: 893– 899.
- Hammer, K.A., Carson, C.F., & Riley, T.V. (1999): Antimicrobial activity of essential oils and other plant extracts. J. Appl. Microbiol., 86: 985–990.
- Hashem, M., Moharam, A.M., Zaired, A.A., & Saleh, F.E.M. (2010): Efficacy of essential oils in the control of cumin root disease by *Fusarium* spp. Crop Prot., 29: 1111– 1117.
- Helal, G.A., Sarhan, M.M., Abu Shahla, A.N., & Abou El-Khair, E.K. (2006): Antimicrobial activity of some essential oils against microorganisms deteriorating fruit juices. *Mycobiology*, 34: 219–229.
- Kalman, B., Abraham, D., Graph, S., Perl-Treves, R., Meller Harel, Y., & Degani, O. (2020): Isolation and identification of *Fusarium* spp., the causal agents of onion (*Allium cepa*) basal rot in Northeastern Israel. *Biology (Basel)*, 9: 69.
- Kaur, L., & Sharma, S.G. (2021). Identification of plant diseases and distinct approaches for their management. *Bull Natl Res Cent.*, 45: 169.
- Kocić-Tanackov, S., Dimić, G., Lević, J., Tanackov, I., & Tuco, D. (2011): Antifungal activities of basil (*Ocimum basilicum* L.) extract on *Fusarium* species. *Afr. J. Biotechnol.*, 10: 10188–10195.
- Köhl, J., Kolnaar, R., & Ravensberg, W.J. (2019): Mode of action of microbial biological control agents against plant diseases: relevance beyond efficacy. *Front. Plant Sci.*, 10: 845.
- Kumar, P., Mishra, S., Kumar, A., & Sharma, A.K. (2016): Antifungal efficacy of plant essential oils against stored grain fungi of *Fusarium* spp. J. Food Sci. Technol., 53: 3725–3734.
- Lahlali, R., Ezrari, S., Radouane, N., Kenfaoui, J., Esmaeel, Q., El Hamss, H., Belabess, Z., & Barka, E.A. (2022): Biological control of plant pathogens: a global perspective. *Microorganisms*, 10: 596.
- Mamgain, A., Roychowdhury, R., & Tah, J. (2013): Alternaria pathogenicity and its strategic controls. *Res. J. Biol.*, 1: 1–9.
- McDonald, B.A., & Stukenbrock, E.H. (2016): Rapid emergence of pathogens in agroecosystems: global threats to agricultural sustainability and food security. *Philos. Trans. R. Soc. B.*, 371: 20160026.
- Nile, A.S., Kwon, Y.D., & Nile, S.H. (2019): Horticultural oils: possible alternatives to chemical pesticides and insecticides. *Environ. Sci. Pollut. Res. Int.*, 26: 21127– 21139.
- Nxumalo, K.A., Aremu, A.O., & Fawole, O.A. (2021): Potentials of medicinal plant extracts as an alternative to synthetic chemicals in postharvest protection and preservation of horticultural crops: a review. *Sustainability*, 13: 5897.
- Omran, S.M., & Esmailzadeh, S. (2009): Comparison of anti-Candida activity of thyme, pennyroyal, and lemon essential oils versus antifungal drugs against Candida species. *Jundishapur J. Microbiol.*, 2: 53–60.
- Pandey, V.N., & Dubey, N.K. (1994): Antifungal potential of leaves and essential oils from higher plants against soil phytopathogens. *Soil Biol. Biochem.*, 26: 1417–1421.
- Peng, Y., Li, S.J., Yan, J., Tang, Y., Cheng, J.P., Gao, A.J., Yao, X., Ruan, J.J., & Xu, B.L. (2021): Research progress on phytopathogenic fungi and their role as biocontrol agents. *Front. Microbiol.*, 12: 670135.

- Perrone, G., Susca, A., Cozzi, G., Ehrlich, K., Varga, J., Frisvad, J.C., Meijer, M., Noonim, P., Mahakarnchanakul, W., & Samson, R.A. (2007): Biodiversity of Aspergillus species in some important agricultural products. *Stud. Mycol.*, 59: 53–66.
- Salameh, P., Waked, M., Baldi, I., Brochard, P., & Saleh, B.A. (2006): Respiratory diseases and pesticide exposure: a case-control study in Lebanon. J. Epidemiol. Community Health, 60: 256–261.
- Santra, H.K., & Banerjee, D. (2020): Natural products as fungicide and their role in crop protection. In J. Singh & A.N. Yadav, eds. Natural Bioactive Products in Sustainable Agriculture, pp. 131–219. Springer Nature Singapore. 307 pp.
- Soliman, K.M., & Badeaa, R.I. (2002): Effect of oil extracted from some medicinal plants on different mycotoxigenic fungi. *Food Chem. Toxicol.*, 40: 1669–1675.
- Stevenson, P.C., Isman, M.B., & Belmain, S.R. (2017): Pesticidal plants in Africa: a global vision of new biological control products from local uses. *Ind. Crops Prod.*, 110: 2– 9.
- Tsegaye, Z., Assefa, F., Tefera, G., Alemu, T., Gizaw, B., & Abatenh, E. (2018): Concept, principle and application of biological control and their role in sustainable plant diseases management strategies. *Int. J. Res. Stud. Biosci.*, 6: 18–34.
- Zheng, S., Chen, B., Qiu, X., Chen, M., Ma, Z., & Yu, X. (2016): Distribution and risk assessment of 82 pesticides in Jiulong River and estuary. *Chemosphere*, 144: 1177– 1192.